ANTIOXIDANTS & REDOX SIGNALING Volume 16, Number 9, 2012 © Mary Ann Liebert, Inc.

DOI: 10.1089/ars.2011.4419

Targeting Mitochondria for Neuroprotection in Parkinson's Disease

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Abstract

Significance: Several genetic causes of familial Parkinson's disease (PD) have now been identified and include mutations of genes encoding mitochondrial proteins. Mitochondrial complex I toxins can induce dopaminergic cell death and produce a parkinsonian state. Importantly, defects of mitochondrial function have been identified in postmortem substantia nigra from pathologically proven cases of PD. Recent Advances: These observations provide compelling evidence to support the notion that mitochondria play an important role in the pathogenesis of PD. Thus, targeting mitochondrial function to delay or prevent neuronal cell death would represent a logical means to modify the course of this disease. Several attempts have already been made in this respect, and have been tested in clinical trial. Critical Issues: To date, there is no unequivocal evidence for an effective intervention to slow the disease. However, several novel mitochondrial targets are now emerging, including the potential to manipulate the mitochondrial pool to maintain function via biogenesis and mitophagy. Future Directions: This development in drug targets needs to be supported by a parallel improvement in clinical trial design to be able to detect a neuroprotective or disease-modifying effect over a reasonable time scale. Antioxid. Redox Signal. 16, 965–973.

Introduction

THERE IS EVIDENCE from a comprehensive range of sources ▲ that mitochondria play a significant role in the pathogenesis of Parkinson's disease (PD). This topic has been the subject of recent reviews (57, 51) and will be covered by other articles in this Forum. This contribution may encompass both primary and secondary roles, as reflected by the genetic causes of some familial forms of PD (Table 1), and by the range of mitochondrial defects identified in postmortem PD brains. An important consideration is whether mitochondria can serve as a useful target to modify function and slow progression in PD. The concept of neuroprotection in PD includes both neuroprotection and neurorestoration (52) (Fig. 1a, b). Targeting mitochondria could potentially include these two effects if dysfunctional neurons were restored to normal activity and death of neurons was delayed or prevented. The interactions of the biochemical pathways and mitochondrial function involved in PD pathogenesis are summarized in Figure 2a and 2b.

This review will discuss attempts to manipulate mitochondrial function in PD that have already been undertaken, or in progress, and potential future targets for consideration.

Respiratory Chain Function and Oxidative Stress

The mitochondrion is a major source of reactive oxygen species by virtue of the reduction of oxygen by the respiratory chain and oxidative phosphorylation (OXPHOS) system. Defects of respiratory chain function, whether inherent or induced by toxins, cause an excess of free radical production. Senescence is associated with accumulating mitochondrial defects including declining mitochondrial OXPHOS function and increasing mitochondrial DNA mutations (30). This is of clear relevance to PD given that it is predominantly an age-related disease. Thus, any mitochondrial defect in PD will develop on the background of mitochondrial dysfunction due to age.

Coenzyme Q10 is an electron carrier and antioxidant, and is an integral part of the respiratory chain. This compound has been used for many years in the treatment of primary mitochondrial disease, although no formal clinical trial of sufficient size has been performed to confirm efficacy (36). In early PD patients, a dose of 1200 mg/day, coenzyme Q10 appeared to slow progression in a pilot study over a period of 16 months (69). Other studies using similar bioavailable levels have either failed to show benefit (71, 79). Additional clinical trials of coenzyme Q10 in early PD are underway and due to report in 2012. Coenzyme Q10 has been used in other

Acronym	Mode of inheritance	Locus	Gene/protein	Main clinical features
PARK1/	Autosomal	4q21-q23	SNCA	Early onset parkinsonism
PARK4	dominant	1 1	(alpha-synuclein)	$(\sim 40 \text{ years})$, dementia
PARK5	Autosomal	4p14	<i>UCHL1</i> (ubiquitin	Typical PD
	dominant	•	carboxyterminal hydrolase 1)	Very rare
PARK8	Autosomal dominant	12q12	LRRK2 (leucine-rich repeat kinase 2)	Typical PD
PARK13	Probably	2p12	Omi/HtrA2	Typical PD
	autosomal dominant	1		Very rare
PARK2	Autosomal recessive	6q25.2-q27	Parkin	Early onset (usually <30 years), rarely juvenile, slow progression
PARK6	Autosomal recessive	1p35-p36	PINK1 PTEN-induced putative kinase 1	Early onset (<40 years), slow progression, psychiatric features common
PARK7	Autosomal recessive	1p36	Dj-1	Early onset (\sim 30–40 years), rarely juvenile
PARK9	Autosomal recessive	1p36	ATP13A2	Early onset, dementia, pyramidal features supranuclear gaze palsy
Not assigned	Probably autosomal dominant	2q22-q23	NR4A2/Nurr1	Typical PD

TABLE 1. GENETIC CAUSES OF MONOGENIC FAMILIAL PARKINSON'S DISEASE

Adapted from Reference 62. PD, Parkinson's disease.

neurodegenerative disorders including Huntington's disease (HD) and Friedreich ataxia. In HD it showed a trend to slowing progression, but did not reach significance (78). In Friedreich's ataxia, a combination of coenzyme Q10 and vitamin E improved neurological progression compared with 4-year cross-sectional natural progression data (19) and low dose was as effective as high dose therapy in improving function in those with coexisting coenzyme Q deficiency (11).

Creatine has also been investigated in PD and at a dose of 10 g daily, was well tolerated, and satisfied the predetermined criterion for nonfutility based on time to requirement for symptomatic therapy for 66 early PD patients (45). Another blinded placebo controlled study of 2 g daily for 6 months then 4 g daily for 18 months in 31 PD patients compared with 17 placebo showed no significant differences compared with placebo (3).

For an agent that is to have an effect upon mitochondrial function, adequate penetration into both the central nervous system and into mitochondria is essential. There are attempts to produce modified agents with either a positive charge that can be concentrated into mitochondria or a mitochondrial targeting sequence. Mito-Q is an example of the former and in vitro studies have demonstrated good mitochondrial penetration and activity (37). This compound is under evaluation in clinical trial. An interesting paradigm for brain penetration and mitochondrial concentration is 1-methyl-4-phenyl 1,2,3,6 tetrahydropyridine (MPTP). This toxin is widely distributed after intravenous administration and has good brain penetration. It is converted to 1-methyl 4-phenylpyridinium (MPP+) by monoamine oxidase (MAO) in glia. MPP+ is actively concentrated into mitochondria where it acts as a complex I inhibitor and free radical generator (10, 41) and inducer of a parkinsonian phenotype (29).

Apoptosis

The mitochondrion is an important mediator of intracellular signals for apoptosis (23, 81, 83). In part this depends on a sequence of mitochondrial depolarization and loss of membrane potential with the release of cytochrome c and the activation of a series of proapoptotic molecules (caspase cascade). Impaired function of OXPHOS or increased oxidative stress could act to lower the membrane potential and thus lower the threshold for apoptosis. A mutation in the mitochondrial-associated apoptosis initiating factor has recently been associated with X-linked infantile onset mitochondrial encephalomyopathy, partially responsive to riboflavin (18).

There are many potential therapeutic targets in the mitochondrial apoptosis cascade. The principle would be to inhibit apoptosis and so prevent cell death. However, such a strategy would have to be specific for neurons and would raise concerns regarding tumorogenesis. Furthermore, simply preventing damaged cells from dying may not restore function to the pathways involved. Despite these concerns, several agents have demonstrated antiapoptotic activity and have been considered for use in slowing progression in PD. Among these are selegiline and rasagiline, two irreversible inhibitors of MAO, with specificity for MAO-B. MAO is responsible for the breakdown of dopamine and its inhibition therefore increases synaptic dopamine levels and reuptake into presynaptic vesicles.

Selegiline

A number of studies have demonstrated the neuroprotective properties of selegiline (L-deprenyl) and its metabolite, desmethylselegiline (DMS), in rodents and primates (17, 20, 26, 35, 74). Early studies by Heikkila and colleagues

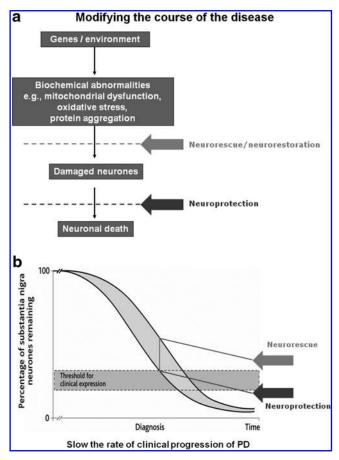


FIG. 1. (a and b) Neurorescue and neuroprotection in Parkinson's disease. Effective neurorescue at diagnosis will restore damaged neurons that are at risk of death (shaded area between curves) to normal function, and age-related loss will probably be attenuated with continuing treatment. Neuroprotection will prevent further neuronal loss other than by attenuated age-related loss (Adapted from Ref. 52).

established that MAO-B inhibitors, including selegiline, could block MPTP-induced degeneration of the nigrostriatal pathway in mice by preventing the conversion of MPTP to its toxic metabolite, MPP+ (20). Tatton and Greenwood later showed that selegiline could increase survival of dopamine neurons in the substantia nigra in mice through a mechanism independent of blockade of MPTP conversion to MPP+ (74). Subsequent studies by Matsubara and colleagues showed that selegiline reduced lactate accumulation in the striatum following MPP+ and suggested that this reflected the improved function of OXPHOS (35).

Selegiline and DMS protected SH-SY5Y cells against apoptosis induced by peroxynitrate, a reactive oxygen species generated from *N*-morpholino sydnonimine (SIN-1), and suggested that protection was independent of MAO-B activity, since SH-SY5Y cells contain only MAO-A (17, 26, 33). It was suggested that (—)-deprenyl exerted antiapoptotic effects through the modulation of gene expression and induction of new protein synthesis rather than MAO inhibition (75, 76). Selegiline and DMS prevent neurotoxin and trophic factor withdrawal-induced cell death by virtue of their ability to induce antiapoptotic molecules, such as Bcl-xL, Bcl-2, glutathione, and superoxide dismutases 1 and 2 (73, 77). These

molecules maintain closure of the mitochondrial permeability transition pore (PTP), whose opening is an important component of the signaling process leading to apoptosis (32, 67). Selegiline/DMS also exert antiapoptotic effects by down-regulating proapoptotic molecules, including c-Jun (which activates the caspase cascade), nitrous oxide synthase, and Bax (which promotes apoptosis by inducing PTP opening), and by preventing nuclear translocation of glyceraldehyde-3-phosphate dehydrogenase, an enzyme whose activation is involved in the early stages of apoptosis (32, 38, 67). Thus, it now appears that (—)-deprenyl-related propargylamines exert antiapoptotic effects *via* alteration of proteins involved in oxidative damage, mitochondrial membrane permeability, or apoptotic signaling pathways (77).

Several clinical studies have sought to evaluate the potential for selegiline to have a neuroprotective or disease modifying effect. The Deprenyl And Tocopherol Antioxidative Therapy Of Parkinsonism (DATATOP) study, a double-blind, multicenter, placebo-controlled clinical trial was undertaken in early PD (80). The clinical trial design combined selegiline with α-tocopherol. The DATATOP trial specifically evaluated whether chronic administration of deprenyl (10 mg/day) and/or tocopherol (2000 IU/day) to patients with early, untreated PD (duration of illness ≤5 years) would extend the time until sufficient disability developed to require L-dopa therapy (primary end point) (80). Analysis at 12±5 months of the study's start revealed the emergence of a major treatment effect: 176 of 401 subjects in the placebo group but only 97 of 399 subjects in the deprenyl groups had reached the primary end point (development of sufficient disability to require L-dopa therapy) (31). No beneficial effects of tocopherol were detected. The results suggested that chronic administration of selegiline (deprenyl) and/or tocopherol in otherwise untreated PD patients could extend the time until disability required therapy with L-dopa. The DATATOP results are now generally attributed to symptomatic benefits of selegiline (31, 59).

Rasagiline

Rasagiline is up to 15 times more potent than selegiline as an MAOB inhibitor in animal models *in vivo* and *in vitro* (2, 86). Rasagiline is metabolized to aminoindan *via* the hepatic cytochrome *P450* (*CYP*) isozyme *CYP1A2* (8). While some reports suggest that the aminoindan metabolite of rasagiline has neuroprotective activity, *L*-methamphetamine—the major metabolite of selegiline—has neurotoxic activity *in vitro* and blocks the neuroprotective action of selegiline and rasagiline (2).

In vitro studies support structure-activity analysis that suggests the antiapoptotic, neuroprotective activity of rasagiline resides in the propargyl moiety and is not related to MAO inhibition. TVP1022, the S-enantiomer of rasagiline, has 1000-fold weaker MAO inhibitory activity but exhibits similar neuroprotective effects *in vitro* (34, 84, 85). The neuroprotective activity of rasagiline has been demonstrated *in vitro* against a variety of neurotoxic insults, including the nitric oxide donor SIN-1, as well as glutamate, 6-hydroxydopamine (6-OHDA), MPTP, β -amyloid, 1,2,3,4-tetrahydroisoquinoline, and serum and growth factor deprivation (32).

A number of *in vitro* studies have suggested several molecular mechanisms by which rasagiline prevents apoptosis (reviewed in 55, 58). Rasagiline significantly reduced cell

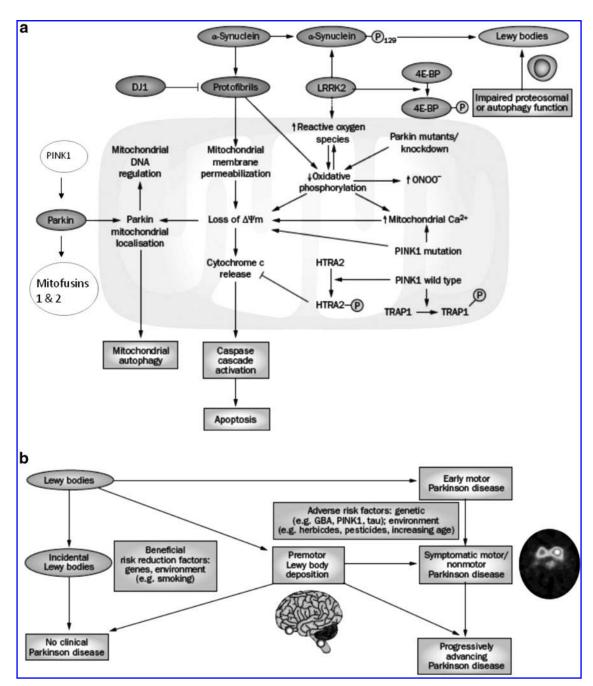


FIG. 2. (a and b) Interconnecting pathways of Parkinson's disease pathogenesis. The mitochondrion plays an important role as a common effector pathway for multiple genetic causes of PD (a). PINK1, DJ1, and parkin proteins have important roles in mitochondrial function, mitochondrial turnover, and free radical metabolism. The mitochondrion in turn is crucial in energy metabolism and as a mediator of the apoptosis pathway. Alpha synuclein has recently been shown to influence oxidative phosphorylation (OXPHOS) and its phosphorylation at position 129 is indirectly influenced by LRRK2 protein. LRRK2 protein may in part have a mitochondrial location (probably outer membrane). The formation of Lewy bodies is central to PD pathology (b), but their role (protective/destructive) is not accurately defined. Lewy bodies maybe deposited without progression to PD (incidental Lewy body "disease"), in extra-nigral sites in patients with premotor disease (Braak stages I and II) or in the nigra associated with early PD. The evolution or transformation between these groups may be driven by several genetic or environmental factors that influence the pathways leading to Lewy body formation and neuronal dysfunction (Adapted from Ref. 64).

death following serum deprivation, and prevented the appearance of cleaved forms of caspase-3 and the caspase substrate poly(ADP-ribose) polymerase (PARP). Results showed that GF109203×, a broad-spectrum protein kinase C (PKC) inhibitor, markedly reversed rasagiline's suppressive effect

on the cleavage and activation of caspase-3 and PARP, suggesting that the PKC pathway mediates neuroprotection by rasagiline. Reverse transcriptase-polymerase chain reaction analysis revealed that treatment of PC12 cells with rasagiline for 24 h significantly increased expression of the PKC

isoenzymes PKC- α and PKC- ϵ and antiapoptotic Bcl-2 family members Bcl-xL and Bcl-w, while decreasing the proapoptotic Bcl-2 family member, Bad. These authors suggested that the activation of PKC in association with the Bcl-2 protein family mediates the neuroprotective activity of rasagiline.

The potential for rasagiline to modify the progression of PD was assessed in the Attenuation of Disease progression with Azilect Given Once-daily (ADAGIO) study. This was an 18-month, double-blind, placebo-controlled, multicenter trial that used a delayed-start design (44). The delayed-start design can, with certain assumptions, define a difference between a pure symptomatic effect and one that maybe associated with modification of the progression of the disease (51, 53). Thus, in ADAGIO, 1176 patients with early PD were randomized to 1 or 2 mg of rasagiline or placebo and maintained blind for 9 months at which point patients were re-randomized to take 1 or 2 mg of rasagiline. In effect this involved those patients already taking a dose of active drug continuing on this, while those on placebo were transferred to one of the two doses of rasagiline.

The primary analysis comprised three hierarchical end points based on the change from baseline in the total Unified Parkinson's Disease Rating Scale (UPDRS) score. In the first, the rate of progression after week 12 had to be significantly slower during the first 9 months in the early start group. For the second, there had to be a significant difference at 18 months in UPDRS scores between the early and late starters. Finally, the rate of progression of UPDRS in months 9 to 18 should be at least parallel and not converging. The 1 mg dose of rasagiline met all three end points (Fig. 3); the 2 mg dose met the first, but failed the second and therefore failed the hierarchical analysis.

The mechanism by which 1 mg rasagiline caused its effect is not revealed by the ADAGIO trial. Indeed, there are several interpretations (54, 60, 61). One mechanism may be through the upregulation of protective and downregulation of harmful compensatory mechanisms that have developed over the long prodrome of PD and maintained normal motor function despite falling dopamine levels (56). Alternatively, rasagiline may be exerting a true protective effect, in line with the laboratory data presented previously. Nevertheless, a symp-

tomatic action that accumulates over a period greater than 9 months, although unlikely, cannot be excluded (63).

Post hoc analysis of the ADAGIO study has shown that the only significant difference between the early and late starters was in the activities of daily living scores, with the 1 mg dose (49). This also showed that those patients with the lower UPDRS scores at baseline progressed slower than those with higher scores, and that the younger patients tended to progress faster (65).

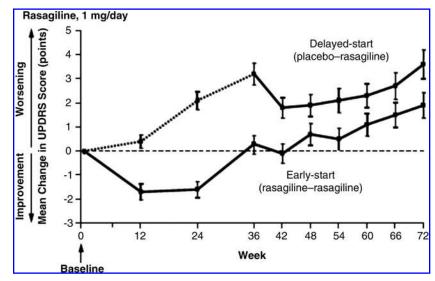
Other inhibitors of apoptosis have also come to clinical trial, but have not demonstrated an ability to slow progression of PD (28, 42).

Autophagy

Mitochondria may divide or bud (fission) or fuse, and this fission–fusion process is regulated by several signaling proteins including mitofusins 1 and 2 that mediate fusion of mitochondrial outer membranes, and optic atrophy protein 1, a dynamin-like GTPase involved in fusion of the inner membranes. Dynamin-1–like protein 1 and fission 1 homologue regulate mitochondrial fission (9).

The fission-fusion process maintains the effectiveness, or quality, of cell mitochondria (6, 7, 12). Defective mitochondria, as identified by a fall in mitochondrial potential, may recover and return to the mitochondrial pool by fusion, or be destroyed by autophagy. The process of mitochondrial destruction by autophagy is referred to as "mitophagy." Parkin and PINK1 proteins have recently been identified as playing an important role in the fission-fusion and mitophagy pathways (13, 47, 66). Parkin translocates from the cytosol to the mitochondrion in response to a fall in mitochondrial membrane potential (39). Recent data suggest that this is preceded by phosphorylation of parkin by PINK1 (13, 25, 39, 47), although other studies have failed to replicate this observation (40, 82). Parkin and PINK1 involvement in mitophagy includes the ubiquitination of MFN 1 and 2, and the mitochondrial voltage-gated anion channel by parkin (14, 16). HtrA2 is thought to participate in mitochondrial turnover and has also been identified as a phosphorylation target of PINK1 (46). The relevance of mitophagy to mitochondrial pathology

FIG. 3. Results of rasagiline 1 mg/day delayed-start study. The mean (±SE) change from baseline in the UPDRS score in the efficacy cohort for the second and third primary end points for patients receiving rasagiline at a dose of 1 mg per day. The dashed lines indicate placebo, and the solid lines indicate rasagiline. UPDRS, Unified Parkinson's Disease Rating Scale (Adapted from Ref. 44).



is emphasized by evidence that defects of the mitophagy pathway contribute to PD pathogenesis (15). The phosphorylation of HtRA2 is dependent on PINK1, probably *via* a kinase cascade, rather than as a direct substrate (46). Mutations in the *HtRA2* gene are a possible cause of familial PD (5). The mitochondrial chaperone TRAP1 has been shown to be a direct substrate of PINK1 (48). These data suggest that PINK1 might be involved in the degradation of mitochondrial proteins as well as mitochondria as a whole.

Importantly, it now appears that mutations of *parkin* or *PINK1* that cause PD interfere with mitophagy efficiency and result in accumulation of defective mitochondria (14). This can be reversed by upregulation of *parkin* or *PINK1* and the removal of defective mitochondria. The recent demonstration of abnormal expression of autophagy proteins in PD brain has further highlighted the importance of degradation pathways to the pathogenesis of this disease (1). These observations offer intriguing opportunities for the development of future therapies for disorders that involve impaired mitochondrial function. In this context it is of note that upregulation of the translation inhibitor 4E-BP ameliorates the effects of PINK1/parkin mutants in *Drosophila*, and rapamycin, a drug that activates 4E-BP and autophagy, is also protective in these mutants (72).

Mitochondrial Biogenesis

An interesting strategy for reversing the effects of mitochondrial dysfunction might be through the regulation of mitochondrial protein transcription and biogenesis. Peroxisome proliferator-activated receptor γ (PPAR γ) coactivator-1 α (PGC- 1α) is an important regulator of mitochondrial activity (see box) and functions together with SIRT1 to influence genetic programs for mitochondrial biogenesis. SIRT1 is one of the family of situins, known to catalyze NAD+-dependent protein deacetylation, yielding nicotinamide and O-acetyl-ADPribose and to regulate longevity, apoptosis, and DNA repair (4). SIRT1 interacts directly with and deacetylates PGC- 1α and increases PGC-1a activity leading to the induction of gene transcription. PGC-1α and SIRT1 promote adaptation to caloric restriction by regulating the genetic programs for gluconeogenesis and glycolysis (50). Activation of PPARy transactivates target genes with the support of PGC- 1α . Drugs that activate PPARy include rosiglitazone, pioglitazone, and troglitazone currently available for treatment of diabetes mellitus. Resveratrol (RSV) is a natural polyphenolic compound found in the skin of grapes and has been shown significantly to increase SIRT1 activity (21). RSV enhances PGC-1α activity, increasing mitochondrial biogenesis and rendered animals resistant to diet-induced obesity and insulin resistance (27). RSV or bezafibrate, another PGC-1α agonist, has demonstrated protective properties in animal models of PD (22, 24).

There is evidence that PGC-1 α upregulation enhances antioxidant activity and inhibition of this effect enhances cell death (70). A recently identified parkin interacting protein, termed PARIS, downregulates PGC-1 α but this action is itself inhibited by parkin. Thus this represents a direct axis between a cause of familial PD and PGC-1 α (68).

Conclusion

Mitochondria are considered an important component of PD pathogenesis, and as such represent a reasonable target for therapeutic intervention to modify the progression of the disease. Several attempts have been made to enhance mitochondrial OXPHOS, reduce oxidative stress, or decrease mitochondrial-mediated apoptosis. None have proven unequivocally effective. This probably reflects in part the rather nonspecific nature of these targets. Perhaps more promising pathways include those of intervening in mitochondrial biogenesis and mitophagy, that is, in the turnover of mitochondria to ensure a healthy population of organelles. However, the proportion of PD patients in whom mitochondrial dysfunction plays a prominent or primary role remains unknown. Thus the clinical trials quoted previously very likely contained a heterogeneous PD population in terms of etiopathogenesis. A single therapeutic agent designed only to improve mitochondrial function, including apoptosis, may thus not be beneficial in those in whom these play no part. This also reflects the complexity of clinical trial design for neuroprotection: the population studies, the end points selected, and so on (43, 65, 66). Although much work needs to be done in this area, developments in clinical trial design for disease modification must also progress in parallel.

Author Disclosure Statement

The author has received honoraria for educational symposia and advisory boards for Teva-Lundbeck.

References

- Alvarez-Erviti L, Rodriguez-Oroz MC, Cooper JM, Caballero C, Ferrer I, Obeso JA, and Schapira AH. Chaperone-mediated autophagy markers in Parkinson disease brains. *Arch Neurol* 67: 1464–1472, 2010.
- Bar AO, Amit T, and Youdim MB. Contrasting neuroprotective and neurotoxic actions of respective metabolites of anti-Parkinson drugs rasagiline and selegiline. *Neurosci Lett* 355: 169–172, 2004.
- Bender A, Koch W, Elstner M, Schombacher Y, Bender J, Moeschl M, Gekeler F, Muller-Myhsok B, Gasser T, Tatsch K, and Klopstock T. Creatine supplementation in Parkinson disease: a placebo-controlled randomized pilot trial. *Neurology* 67: 1262–1264, 2006.
- 4. Blander G and Guarente L. The Sir2 family of protein deacetylases. *Annu Rev Biochem* 73: 417–435, 2004.
- Bogaerts V, Nuytemans K, Reumers J, Pals P, Engelborghs S, Pickut B, Corsmit E, Peeters K, Schymkowitz J, De Deyn PP, Cras P, Rousseau F, Theuns J, and Van BC. Genetic variability in the mitochondrial serine protease HTRA2 contributes to risk for Parkinson disease. *Hum Mutat* 29: 832–840, 2008.
- Chan DC. Mitochondria: dynamic organelles in disease, aging, and development. Cell 125: 1241–1252, 2006.
- Chen JH, Stoltz DA, Karp PH, Ernst SE, Pezzulo AA, Moninger TO, Rector MV, Reznikov LR, Launspach JL, Chaloner K, Zabner J, and Welsh MJ. Loss of anion transport without increased sodium absorption characterizes newborn porcine cystic fibrosis airway epithelia. *Cell* 143: 911–923, 2010
- Chen JJ and Swope DM. Clinical pharmacology of rasagiline: a novel, second-generation propargylamine for the treatment of Parkinson disease. J Clin Pharmacol 45: 878–894, 2005.
- Cho DH, Nakamura T, and Lipton SA. Mitochondrial dynamics in cell death and neurodegeneration. *Cell Mol Life Sci* 67: 3435–3447, 2010.

- Cleeter MW, Cooper JM, and Schapira AH. Irreversible inhibition of mitochondrial complex I by 1-methyl-4-phenylpyridinium: evidence for free radical involvement. J Neurochem 58: 786–789, 1992.
- 11. Cooper JM, Korlipara LV, Hart PE, Bradley JL, and Schapira AH. Coenzyme Q10 and vitamin E deficiency in Friedreich's ataxia: predictor of efficacy of vitamin E and coenzyme Q10 therapy. *Eur J Neurol* 15: 1371–1379, 2008.
- 12. Dagda RK and Chu CT. Mitochondrial quality control: insights on how Parkinson's disease related genes PINK1, parkin, and Omi/HtrA2 interact to maintain mitochondrial homeostasis. *J Bioenerg Biomembr* 41: 473–479, 2009.
- 13. Deng H, Dodson MW, Huang H, and Guo M. The Parkinson's disease genes pink1 and parkin promote mitochondrial fission and/or inhibit fusion in Drosophila. *Proc Natl Acad Sci U S A* 105: 14503–14508, 2008.
- 14. Gegg ME, Cooper JM, Chau KY, Rojo M, Schapira AH, and Taanman JW. Mitofusin 1 and mitofusin 2 are ubiquitinated in a PINK1/parkin-dependent manner upon induction of mitophagy. *Hum Mol Genet* 19: 4861–4870, 2010.
- Gegg ME and Schapira AH. PINK1-parkin-dependent mitophagy involves ubiquitination of mitofusins 1 and 2: Implications for Parkinson disease pathogenesis. *Autophagy* 7: 243–245, 2011.
- Geisler S, Holmstrom KM, Skujat D, Fiesel FC, Rothfuss OC, Kahle PJ, and Springer W. PINK1/Parkin-mediated mitophagy is dependent on VDAC1 and p62/SQSTM1. Nat Cell Biol 12: 119–131, 2010.
- Gerlach M, Foley P, and Riederer P. The relevance of preclinical studies for the treatment of Parkinson's disease. J Neurol 250 Suppl 1: I31–I34, 2003.
- Ghezzi D, Sevrioukova I, Invernizzi F, Lamperti C, Mora M, D'Adamo P, Novara F, Zuffardi O, Uziel G, and Zeviani M. Severe X-linked mitochondrial encephalomyopathy associated with a mutation in apoptosis-inducing factor. *Am J Hum Genet* 86: 639–649, 2010.
- Hart PE, Lodi R, Rajagopalan B, Bradley JL, Crilley JG, Turner C, Blamire AM, Manners D, Styles P, Schapira AH, and Cooper JM. Antioxidant treatment of patients with Friedreich ataxia: four-year follow-up. *Arch Neurol* 62: 621–626, 2005.
- Heikkila RE, Manzino L, Cabbat FS, and Duvoisin RC. Protection against the dopaminergic neurotoxicity of 1methyl-4-phenyl-1,2,5,6-tetrahydropyridine by monoamine oxidase inhibitors. *Nature* 311: 467–469, 1984.
- Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, Zipkin RE, Chung P, Kisielewski A, Zhang LL, Scherer B, and Sinclair DA. Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. *Nature* 425: 191–196, 2003.
- 22. Jin F, Wu Q, Lu YF, Gong QH, and Shi JS. Neuroprotective effect of resveratrol on 6-OHDA-induced Parkinson's disease in rats. *Eur J Pharmacol* 600: 78–82, 2008.
- Karbowski M. Mitochondria on guard: role of mitochondrial fusion and fission in the regulation of apoptosis. Adv Exp Med Biol 687: 131–142, 2010.
- 24. Khan MM, Ahmad A, Ishrat T, Khan MB, Hoda MN, Khuwaja G, Raza SS, Khan A, Javed H, Vaibhav K, and Islam F. Resveratrol attenuates 6-hydroxydopamine-induced oxidative damage and dopamine depletion in rat model of Parkinson's disease. *Brain Res* 1328: 139–151, 2010.
- Kim Y, Park J, Kim S, Song S, Kwon SK, Lee SH, Kitada T, Kim JM, and Chung J. PINK1 controls mitochondrial localization of Parkin through direct phosphorylation. *Biochem Biophys Res Commun* 377: 975–980, 2008.

- Kupsch A, Sautter J, Gotz ME, Breithaupt W, Schwarz J, Youdim MB, Riederer P, Gerlach M, and Oertel WH. Monoamine oxidase-inhibition and MPTP-induced neurotoxicity in the non-human primate: comparison of rasagiline (TVP 1012) with selegiline. J Neural Transm 108: 985–1009, 2001.
- Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, Messadeq N, Milne J, Lambert P, Elliott P, Geny B, Laakso M, Puigserver P, and Auwerx J. Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell* 127: 1109–1122, 2006.
- Lang AE. Clinical trials of disease-modifying therapies for neurodegenerative diseases: the challenges and the future. *Nat Med* 16: 1223–1226, 2010.
- Langston JW, Ballard P, Tetrud JW, and Irwin I. Chronic Parkinsonism in humans due to a product of meperidineanalog synthesis. Science 219: 979–980, 1983.
- 30. Larsson NG. Somatic mitochondrial DNA mutations in mammalian aging. *Annu Rev Biochem* 79: 683–706, 2010.
- 31. LeWitt PA. Clinical trials of neuroprotection for Parkinson's disease. *Neurology* 63: S23–S31, 2004.
- 32. Mandel S, Weinreb O, Amit T, and Youdim MB. Mechanism of neuroprotective action of the anti-Parkinson drug rasagiline and its derivatives. *Brain Res Brain Res Rev* 48: 379–387, 2005.
- Maruyama W and Naoi M. Neuroprotection by (-)-deprenyl and related compounds. *Mech Ageing Dev* 111: 189–200, 1999.
- 34. Maruyama W, Weinstock M, Youdim MB, Nagai M, and Naoi M. Anti-apoptotic action of anti-Alzheimer drug, TV3326 [(N-propargyl)-(3R)-aminoindan-5-yl]-ethyl methyl carbamate, a novel cholinesterase-monoamine oxidase inhibitor. *Neurosci Lett* 341: 233–236, 2003.
- 35. Matsubara K, Senda T, Uezono T, Awaya T, Ogawa S, Chiba K, Shimizu K, Hayase N, and Kimura K. L-Deprenyl prevents the cell hypoxia induced by dopaminergic neurotoxins, MPP(+) and beta-carbolinium: a microdialysis study in rats. *Neurosci Lett* 302: 65–68, 2001.
- 36. Morgan-Hughes JA, Schapira AH, Cooper JM, and Clark JB. Molecular defects of NADH-ubiquinone oxidoreductase (complex I) in mitochondrial diseases. *J Bioenerg Biomembr* 20: 365–382, 1988.
- 37. Murphy MP and Smith RA. Targeting antioxidants to mitochondria by conjugation to lipophilic cations. *Annu Rev Pharmacol Toxicol* 47: 629–656, 2007.
- Naoi M and Maruyama W. Future of neuroprotection in Parkinson's disease. *Parkinsonism Relat Disord* 8: 139–145, 2001.
- 39. Narendra D, Tanaka A, Suen DF, and Youle RJ. Parkin is recruited selectively to impaired mitochondria and promotes their autophagy. *J Cell Biol* 183: 795–803, 2008.
- Narendra DP, Jin SM, Tanaka A, Suen DF, Gautier CA, Shen J, Cookson MR, and Youle RJ. PINK1 is selectively stabilized on impaired mitochondria to activate Parkin. *PLoS Biol* 8: e1000298, 2010.
- Nicklas WJ, Vyas I, and Heikkila RE. Inhibition of NADHlinked oxidation in brain mitochondria by 1-methyl-4-phenyl-pyridine, a metabolite of the neurotoxin, 1-methyl-4phenyl-1,2,5,6-tetrahydropyridine. *Life Sci* 36: 2503–2508, 1985
- Olanow CW, Schapira AH, LeWitt PA, Kieburtz K, Sauer D, Olivieri G, Pohlmann H, and Hubble J. TCH346 as a neuroprotective drug in Parkinson's disease: a double-blind,

randomised, controlled trial. Lancet Neurol 5: 1013–1020, 2006.

- Olanow CW, Kieburtz K, and Schapira AH. Why have we failed to achieve neuroprotection in Parkinson's disease? *Ann Neurol* 64 Suppl 2: S101–S110, 2008.
- 44. Olanow CW, Rascol O, Hauser R, Feigin PD, Jankovic J, Lang A, Langston W, Melamed E, Poewe W, Stocchi F, and Tolosa E. A double-blind, delayed-start trial of rasagiline in Parkinson's disease. *N Engl J Med* 361: 1268–1278, 2009.
- NINDS NET-PD Investigators. A randomized, double-blind, futility clinical trial of creatine and minocycline in early Parkinson disease. *Neurology* 66: 664–671, 2006.
- 46. Plun-Favreau H, Klupsch K, Moisoi N, Gandhi S, Kjaer S, Frith D, Harvey K, Deas E, Harvey RJ, McDonald N, Wood NW, Martins LM, and Downward J. The mitochondrial protease HtrA2 is regulated by Parkinson's disease-associated kinase PINK1. Nat Cell Biol 9: 1243–1252, 2007.
- 47. Poole AC, Thomas RE, Andrews LA, McBride HM, Whitworth AJ, and Pallanck LJ. The PINK1/Parkin pathway regulates mitochondrial morphology. *Proc Natl Acad Sci U S A* 105: 1638–1643, 2008.
- 48. Pridgeon JW, Olzmann JA, Chin LS, and Li L. PINK1 protects against oxidative stress by phosphorylating mitochondrial chaperone TRAP1. *PLoS Biol* 5: e172, 2007.
- 49. Rascol O, Fitzer-Attas CJ, Hauser R, Jankovic J, Lang A, Langston JW, Melamed E, Poewe W, Stocchi F, Tolosa E, Eyal E, Weiss YM, and Olanow CW. A double-blind, delayed-start trial of rasagiline in Parkinson's disease (the ADAGIO study): prespecified and post-hoc analyses of the need for additional therapies, changes in UPDRS scores, and non-motor outcomes. Lancet Neurol 10: 415–423, 2011.
- Rodgers JT, Lerin C, Haas W, Gygi SP, Spiegelman BM, and Puigserver P. Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature* 434: 113–118, 2005.
- 51. Schapira AH. Mitochondrial diseases. Lancet [In press]: 2012.
- 52. Schapira AH. Science, medicine, and the future: Parkinson's disease. *BMJ* 318: 311–314, 1999.
- 53. Schapira AH. Etiology of Parkinson's disease. *Neurology* 66: S10–S23, 2006.
- 54. Schapira AH, Bezard E, Brotchie J, Calon F, Collingridge GL, Ferger B, Hengerer B, Hirsch E, Jenner P, Le NN, Obeso JA, Schwarzschild MA, Spampinato U, and Davidai G. Novel pharmacological targets for the treatment of Parkinson's disease. Nat Rev Drug Discov 5: 845–854, 2006.
- 55. Schapira AH. The use of rasagiline in Parkinson's disease. *J Neural Transm Suppl* 157–161, 2006.
- 56. Schapira AH and Obeso J. Timing of treatment initiation in Parkinson's disease: a need for reappraisal? *Ann Neurol* 59: 559–562, 2006.
- 57. Schapira AH. Mitochondria in the aetiology and pathogenesis of Parkinson's disease. *Lancet Neurol* 7: 97–109, 2008.
- 58. Schapira AH. Rasagiline in neurodegeneration. *Exp Neurol* 212: 255–257, 2008.
- Schapira AH. Molecular and clinical pathways to neuroprotection of dopaminergic drugs in Parkinson disease. Neurology 72: S44–S50, 2009.
- 60. Schapira AH, Emre M, Jenner P, and Poewe W. Levodopa in the treatment of Parkinson's disease. *Eur J Neurol* 16: 982–989, 2009.
- 61. Schapira AH. Neurobiology and treatment of Parkinson's disease. *Trends Pharmacol Sci* 30: 41–47, 2009.

62. Schapira AH, Agid Y, Barone P, Jenner P, Lemke MR, Poewe W, Rascol O, Reichmann H, and Tolosa E. Perspectives on recent advances in the understanding and treatment of Parkinson's disease. *Eur J Neurol* 16: 1090–1099, 2009.

- 63. Schapira AH. Movement disorders: advances in cause and treatment. *Lancet Neurol* 9: 6–7, 2010.
- 64. Schapira AH and Tolosa E. Molecular and clinical prodrome of Parkinson disease: implications for treatment. *Nat Rev Neurol* 6: 309–317, 2010.
- Schapira AH and Schrag A. Parkinson disease: Parkinson disease clinical subtypes and their implications. *Nat Rev Neurol* 7: 247–248, 2011.
- 66. Schapira AH. Challenges to the development of disease-modifying therapies in Parkinson's disease. *Eur J Neurol* 18 Suppl 1: 16–21, 2011.
- 67. Sharma SK, Carlson EC, and Ebadi M. Neuroprotective actions of Selegiline in inhibiting 1-methyl, 4-phenyl, pyridinium ion (MPP+)-induced apoptosis in SK-N-SH neurons. *J Neurocytol* 32: 329–343, 2003.
- Shin JH, Ko HS, Kang H, Lee Y, Lee YI, Pletinkova O, Troconso JC, Dawson VL, and Dawson TM. PARIS (ZNF746) repression of PGC-1alpha contributes to neurodegeneration in Parkinson's disease. Cell 144: 689–702, 2011.
- 69. Shults CW, Oakes D, Kieburtz K, Beal MF, Haas R, Plumb S, Juncos JL, Nutt J, Shoulson I, Carter J, Kompoliti K, Perlmutter JS, Reich S, Stern M, Watts RL, Kurlan R, Molho E, Harrison M, and Lew M. Effects of coenzyme Q10 in early Parkinson disease: evidence of slowing of the functional decline. *Arch Neurol* 59: 1541–1550, 2002.
- St-Pierre J, Drori S, Uldry M, Silvaggi JM, Rhee J, Jager S, Handschin C, Zheng K, Lin J, Yang W, Simon DK, Bachoo R, and Spiegelman BM. Suppression of reactive oxygen species and neurodegeneration by the PGC-1 transcriptional coactivators. *Cell* 127: 397–408, 2006.
- 71. Storch A, Jost WH, Vieregge P, Spiegel J, Greulich W, Durner J, Muller T, Kupsch A, Henningsen H, Oertel WH, Fuchs G, Kuhn W, Niklowitz P, Koch R, Herting B, and Reichmann H. Randomized, double-blind, placebo-controlled trial on symptomatic effects of coenzyme Q(10) in Parkinson disease. *Arch Neurol* 64: 938–944, 2007.
- 72. Tain LS, Mortiboys H, Tao RN, Ziviani E, Bandmann O, and Whitworth AJ. Rapamycin activation of 4E-BP prevents parkinsonian dopaminergic neuron loss. *Nat Neurosci* 12: 1129–1135, 2009.
- 73. Tatton W, Chalmers-Redman R, and Tatton N. Neuroprotection by deprenyl and other propargylamines: glyceral-dehyde-3-phosphate dehydrogenase rather than monoamine oxidase B. *J Neural Transm* 110: 509–515, 2003.
- 74. Tatton WG and Greenwood CE. Rescue of dying neurons: a new action for deprenyl in MPTP parkinsonism. *J Neurosci Res* 30: 666–672, 1991.
- 75. Tatton WG, Ju WY, Holland DP, Tai C, and Kwan M. (-)-Deprenyl reduces PC12 cell apoptosis by inducing new protein synthesis. *J Neurochem* 63: 1572–1575, 1994.
- Tatton WG and Chalmers-Redman RM. Modulation of gene expression rather than monoamine oxidase inhibition: (-)deprenyl-related compounds in controlling neurodegeneration. Neurology 47: S171–S183, 1996.
- Tatton WG, Chalmers-Redman RM, Ju WJ, Mammen M, Carlile GW, Pong AW, and Tatton NA. Propargylamines induce antiapoptotic new protein synthesis in serum- and nerve growth factor (NGF)-withdrawn, NGF-differentiated PC-12 cells. J Pharmacol Exp Ther 301: 753–764, 2002.

- 78. The Huntington Study Group. A randomized, placebocontrolled trial of coenzyme Q10 and remacemide in Huntington's disease. *Neurology* 57: 397–404, 2001.
- 79. The NINDS BET-PD Investigators. A randomized clinical trial of coenzyme Q10 and GPI-1485 in early Parkinson disease. *Neurology* 68: 20–28, 2007.
- The Parkinson Study Group. Effect of deprenyl on the progression of disability in early Parkinson's disease. The Parkinson Study Group. N Engl J Med 321: 1364–1371, 1989.
- 81. Verdin E, Hirschey MD, Finley LW, and Haigis MC. Sirtuin regulation of mitochondria: energy production, apoptosis, and signaling. *Trends Biochem Sci* 35: 669–675, 2010.
- 82. Vives-Bauza C, Zhou C, Huang Y, Cui M, de Vries RL, Kim J, May J, Tocilescu MA, Liu W, Ko HS, Magrane J, Moore DJ, Dawson VL, Grailhe R, Dawson TM, Li C, Tieu K, and Przedborski S. PINK1-dependent recruitment of Parkin to mitochondria in mitophagy. *Proc Natl Acad Sci U S A* 107: 378–383, 2010.
- 83. Wang C and Youle RJ. The role of mitochondria in apoptosis. *Annu Rev Genet* 43: 95–118, 2009.
- 84. Youdim MB, Wadia A, Tatton W, and Weinstock M. The anti-Parkinson drug rasagiline and its cholinesterase inhibitor derivatives exert neuroprotection unrelated to MAO inhibition in cell culture and *in vivo*. *Ann N Y Acad Sci* 939: 450–458, 2001.
- 85. Youdim MB and Weinstock M. Molecular basis of neuroprotective activities of rasagiline and the anti-Alzheimer drug TV3326 [(N-propargyl-(3R)aminoindan-5-YL)-ethyl methyl carbamate]. *Cell Mol Neurobiol* 21: 555–573, 2001.
- 86. Youdim MB, Gross A, and Finberg JP. Rasagiline [N-propargyl-1R(+)-aminoindan], a selective and potent inhibitor of mitochondrial monoamine oxidase B. *Br J Pharmacol* 132: 500–506, 2001.

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Date of first submission to ARS Central, December 16, 2011; date of acceptance, January 2, 2012.

Abbreviations Used

6-OHDA = 6-hydroxydopamine

ADAGIO = Attenuation of Disease progression with Azilect Given Once-daily

DMS = desmethylselegiline

HD = Huntington's disease

MAO = monoamine oxidase

MPP+ = 1-methyl 4-phenylpyridinium

MPTP = 1-methyl-4-phenyl 1,2,3,6 tetrahydropyridine

OXPHOS = oxidative phosphorylation

PARP = poly(ADP-ribose) polymerase

PD = Parkinson's disease

 $PGC-1\alpha = coactivator-1\alpha$

PKC = protein kinase C

PPAR γ = peroxisome proliferator-activated receptor γ

PTP = permeability transition pore

RSV = Resveratrol

SIN-1 = N-morpholino sydnonimine

UPDRS = Unified Parkinson's Disease Rating Scale

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